

# SPE for analysis of environmental contaminants in water

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# OUTLINE

## Extraction Disks

- ENVI Disks vs. Empore
- Applications

## Carbons

- ENVI-Carb
- ENVI-Carb Plus

## SupelSelect HLB for extraction of PPCPs

# Useful EPA 500 Series (drinking water) Accessories



P001063

Visiprep 5-Port Flask  
Manifold



Sample  
Funnel

PTFE Base/  
Adapter

Flask (order  
separately)

ENVI-Disk Holder



Visiprep Lg. Vol. Sampler



ENVI-Disk Holder Manifold

# Extraction Disks

# ENVI-Disk™ SPE Disk vs. Empore SPE Disk

- **ENVI-Disk SPE Disk**

- SPE Particles embedded in glass fiber matrix
- Available in C18 and C8
- Better flow properties but more resistant to clogging
- More prone to fines

- **Empore SPE Disk**

- SPE particles embedded in a PTFE fiber matrix
  - Available in over 9 x different chemistries
  - Minimal fines but more prone to clogging
    - Clogging addressed via Empore Filter Aid
  - Compatible with ENVI-Disk Holder
- Both products written in dozens of EPA & other environmental methods



# SPE Disks

## 3M Empore™ SPE

Reduced SPE bed mass = Reduced solvent & elution volumes

- Minimizes SPE eluate evaporation time
- Potentially allows for direct injection of the SPE eluate

Dense & uniform extraction medium = NO channeling/voiding

- Efficient mass-transfer kinetics allow for faster flow rates



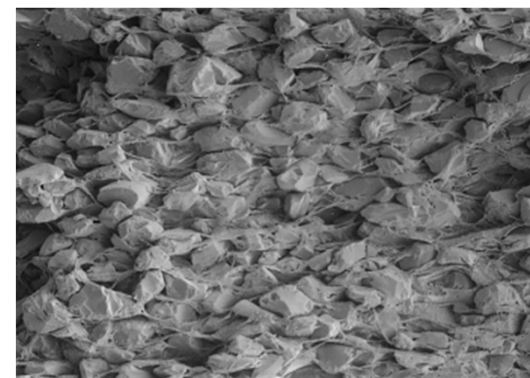
**47 mm Disks for  
Environmental Analysis**

# Empore SPE Disk Technology

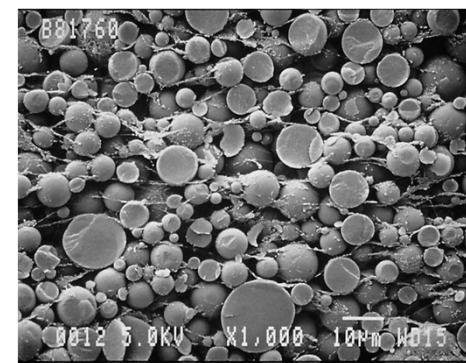
SPE Particles tightly enmeshed in an inert PTFE matrix

- 90% SPE Sorbent; 10% PTFE (by weight)
- Dense particle packing (no void spacing)
- Uniform particle distribution
- Thin membrane, small bed volume
- High Surface Area / No Fines
- Smaller bed weights
- Shorter diffusion paths
- More efficient extractions

**Silica-based**



**Resin-based**



## Official EPA Methods using Empore Disks

- **1664 (Rev. A)** - N-Hexane Extractable Material (HEM; Oil and Grease)
- **506** - Phthalate and Adipate Esters in Drinking Water
- **507** - Nitrogen- and Phosphorus-Containing Pesticides in Water
- **508.1** - Chlorinated Pesticides, Herbicides, and Organohalides in Water
- **512.2** - Chlorinated Acids in Water
- **525.2** - Organic Compounds in Drinking Water
- **549.1** - Diquat and Paraquat in Drinking Water
- **550.1** - Polycyclic Aromatic Hydrocarbons in Drinking Water
- **552.1** - Haloacetic Acids and Dalapon in Drinking Water
- **553** - Benzidines and Nitrogen-Containing Pesticides in Water
- **1613 (Rev.B)** - Tetra- Through Octa-Chlorinated Dioxins and Furans by Isotope Dilution e.g. in water
- **SW846 Method 3535** - Test Methods for TCLP Leachates
- **QTM - Aqueous Phases Quick Turnaround Methods**
  - PAH
  - Phenols
  - Pesticides & PCBs



# Tips to get good performance from extraction disks

## Conditioning

- Prepares sorbent to interact with analytes
- Critical to good recovery and reproducibility
- Do not allow disk to go dry

## Extraction

- Flow rate is not critical to recovery
- After sample, remove water by pulling vacuum

## Elution

- Use multiple aliquots of elution solvent
- Use first aliquot to rinse out sample container
- Soak the disk for one minute with elution solvent prior to pulling vacuum

# Application: Polynuclear Aromatic Hydrocarbons

Method: US EPA 550.1

Disk: 47 mm Empore or ENVI-C18

1. Add 5 mL methanol and IS (if used) to 1 L water sample.
2. Wash disk w/5 mL methylene chloride.
3. Condition disk w/ 5 mL methanol followed by 5 mL dei. water.
4. Process sample through disk at a flow rate of 100 mL/min.
5. Elute the disk with 5 mL acetonitrile and 5 X 5 mL methylene chloride
6. Dry eluate through 3 gm anhydrous sodium sulfate.
7. Concentrate to 0.5 mL and analyze by HPLC

**Column:** Ascentis Express C18, 15 cm x 2.1 mm ID

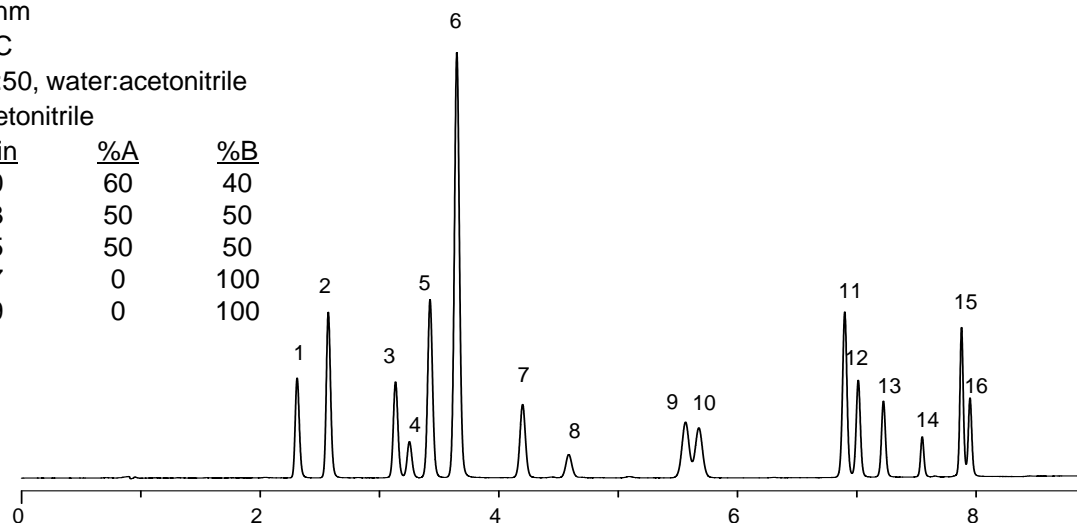
**Detection:** UV, 250 nm

**Temperature:** 35 ° C

**Mobile Phase A:** 50:50, water:acetonitrile

**Mobile Phase B:** acetonitrile

| Gradient: | min | %A | %B  |
|-----------|-----|----|-----|
|           | 0   | 60 | 40  |
|           | 3   | 50 | 50  |
|           | 5   | 50 | 50  |
|           | 7   | 0  | 100 |
|           | 9   | 0  | 100 |



1. Naphthalene
2. Acenaphthylene
3. Fluorene
4. Acenaphthene
5. Phenanthrene
6. Anthracene
7. Fluoranthene
8. Pyrene
9. Chrysene
10. Benzo[a]anthracene
11. Benzo[b]fluoranthene
12. Benzo[k]fluoranthene
13. Benzo[a]pyrene
14. Dibenzo[a,h]anthracene
15. Indeno[1,2,3-cd]pyrene
16. Benzo[g,h,i]perylene

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# Application: Paraquat and Diquat from Water

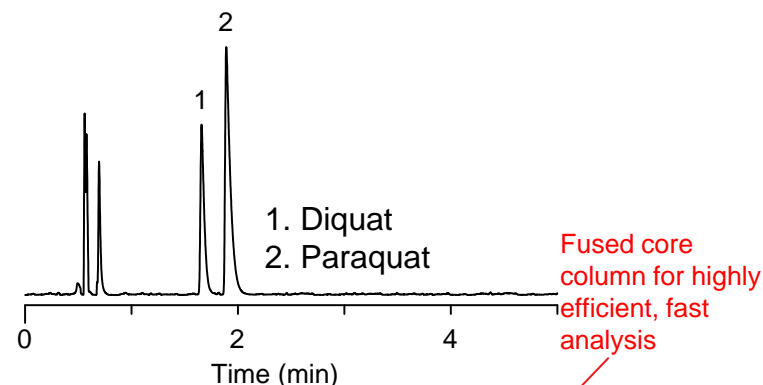
Method: US EPA 549.2

Disk: 47 mm Empore or ENVI-C8

1. Adjust 250 mL water sample to pH 7-9 using 10% NaOH or 10% HCl
2. Condition the disk as follows:
  1. 10 mL methanol
  2. 2 x 10 mL dei. water
  3. 10 mL conditionin soln. A\*
  4. 2x 10 mL dei. water
  5. 20 mL conditioning soln. B\*
3. Process sample through disk at a flow rate of 100 mL/min.
  4. Add 1 mL methanol to disk; soak for 1 min.
  5. Add 4 mL eluting solution\* to disk, apply slight vacuum, and soak disk for 1 min.
  6. Add 4 mL eluting solution\* to disk and draw completely through.
  7. (Add ion pair reagent concentrate to eluate, adjust final volume (if necessary) and analyze by HPLC/UV)

elution

\*see method 549.2 for composition of solution



Column : Ascentis Express HILIC, 10 cm x 2.1 mm I.D., 2.7  $\mu$ m particles (53939-U)  
Mobile Phase: 20:80; 200 mM TFA NH<sub>3</sub>: acetonitrile  
Temperature: 60 °C  
Flow Rate: 0.4 mL/min  
Detection: UV at 308 (diquat) and 257 (paraquat) nm  
Injection Volume: 1  $\mu$ L

**Carbon SPE**  
**ENVI-Carb™ and ENVI-Carb™ Plus**

# Carbon SPE

## ENVI-Carb™

- Predominately GCB (graphitized carbon black)
- Unique Selectivity & Ideal for polar compounds
- Non-porous (adsorption) = faster flow rates
- Example Applications
  - Chloroacetanilide and Chloroacetamide Herbicide Degradents (535.1)
  - Non-volatile pesticides (carbamate & thiourea) in drinking water
  - BNA Pesticides in ground water
  - Acidic herbicides in drinking water (515.2)
  - Nitrosamines in drinking water (521)

| ENVI-Carb Adsorbent                | C8- & C18-Modified Silica              |
|------------------------------------|--|
| graphitized carbon black           | silane phase-modified silica gel       |
| hydrophobic                        | hydrophobic                            |
| irregular 40-100µm particles       | irregular 40-60µm particles            |
| nonporous                          | porous (60-300Å)                       |
| surface area: 100m <sup>2</sup> /g | surface area: 400-600m <sup>2</sup> /g |

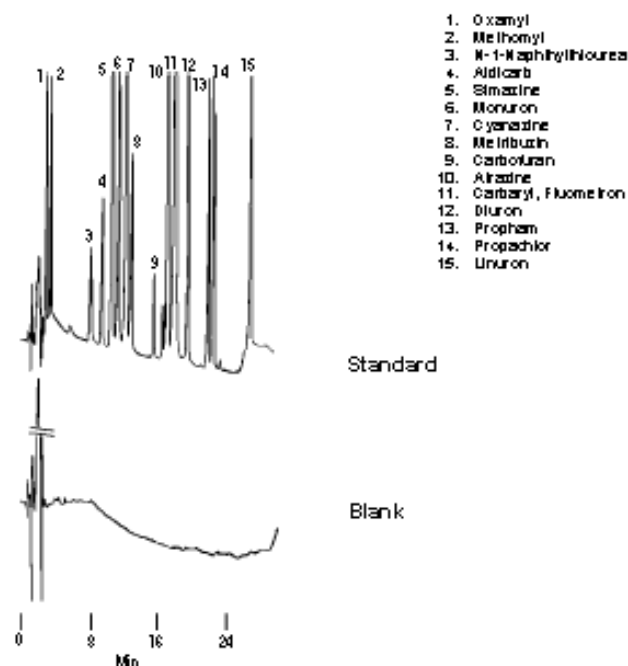
# Application: Use of Envi-Carb for pesticides in water

## Pesticide Extraction from Drinking Water, Using ENVI-Carb Solid Phase Extraction Tubes

SPE Tube:  
Supelclean ENVI-Carb, 3mL/0.25g packing  
Tube Conditioning:  
5mL methylene chloride: methanol (80:20)  
1mL methanol  
10mL 2% acetic acid in water  
Draws solutions through the packing bed consecutively.  
Keep packing bed wet until sample is added.  
Sample Addition:  
100mL 1 liter drinking water (pesticide concentration: 10-50 µg/L)  
Draws sample through packing at a rate of 5mL/min.  
Accelerated rates do not have an adverse effect on recovery.  
Drying:  
1 minute, vacuum suction  
Sample Elution (base-neutral fraction):  
0.8-1mL methanol  
2x 3.5mL methylene chloride: methanol (80:20)  
Dry eluate under gentle nitrogen purge in a room temperature  
water bath to approximately 400-500 µL. For best recovery,  
wash inside wall of recovery vial with methanol,  
again dry to 400-500 µL.  
Reconstitutes samples to a constant volume of 1 mL, using methanol.  
Inject 20 µL for HPLC analysis. Analysis can be automated.

## HPLC Analysis

Column: SUPELCOSIL LC-18-DB, 25cm x 4.6mm (5µm particles)  
(with Supelguard™ LC-18-DB guard column)  
Cat. No.: 58355-U  
Mobile Phase: A - water:acetonitrile, 90:10, B - acetonitrile  
20% B (5 min) to 70% B in 30 min  
Flow Rate: 1.5mL/min  
Det.: UV, 220nm  
Inj.: 20µL extract (see Table 2)



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## Pesticide recovery using ENVI-Carb

| Analyte**  | Solid Phase Extraction                   |  |
|------------|--|--|
|            | ENVI-Carb Tubes                          | C8/C18 Silica*                           |
|            | n=5<br>Recovery (% ± standard deviation) | n=4<br>Recovery (% ± standard deviation) |
| Oxamyl     | 111 ±9.6                                 | 28 ±46                                   |
| Methomyl   | 105 ±5.0                                 | 25 ±44                                   |
| Carbofuran | 106 ±6.2                                 | 97 ±3.8                                  |
| Fluometron | 106 ±5.7                                 | 96 ±4.1                                  |
| Monuron    | 99 ±3.2                                  | 98 ±6.2                                  |
| Metribuzin | 97 ±3.9                                  | 43 ±25                                   |
| Carbaryl   | 97 ±3.5                                  | 90 ±12                                   |
| Propham    | 95 ±3.2                                  | 76 ±23                                   |
| Propachlor | 96 ±3.8                                  | 90 ±7.5                                  |
| Aldicarb   | 96 ±3.5                                  | 86 ±14                                   |
| Cyanazine  | 90 ±5.4                                  | 99 ±10                                   |
| Atrazine   | 89 ±5.7                                  | 74 ±14                                   |
| Diuron     | 88 ±5.7                                  | 91 ±7.8                                  |
| Linuron    | 88 ±5.4                                  | 94 ±4.1                                  |

Low recoveries for polar pesticides on C18/C8

\*\*100mL water samples, 10-50µg/liter (ENVI-Carb extracts) or 20-500µg/liter (C8/C18 silica), HPLC/UV (ENVI-Carb extracts) or HPLC/MS (C8/C18 silica) analysis.

\*Data from Bellar & Budde, *Anal. Chem.*, 60: 2076-2083 (1988).

# Application: Use of ENVI-Carb for extraction of acidic herbicides

## Extraction procedure (automated using Zymark)

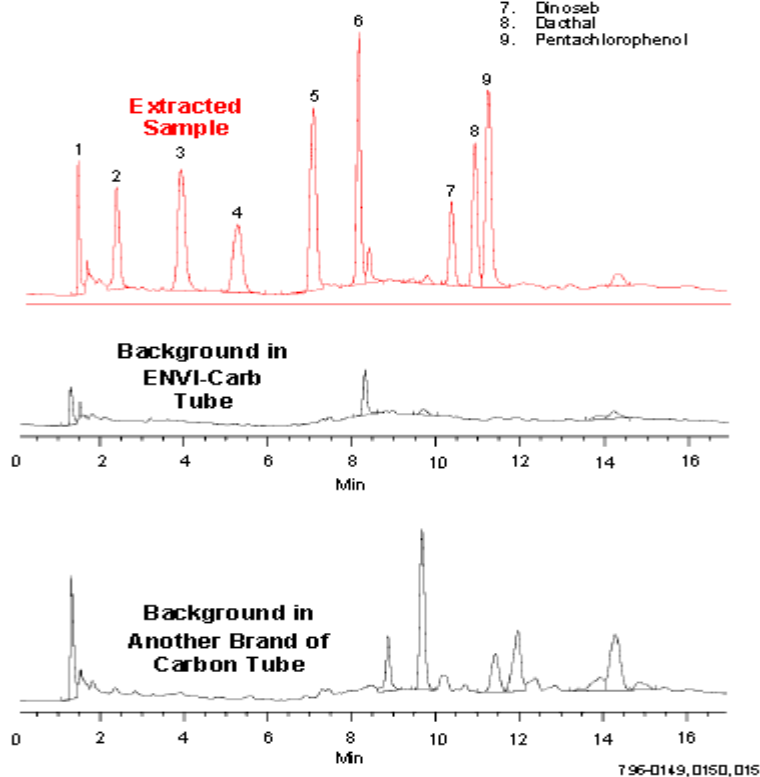
1. Condition 6 mL/250 mg ENVI-Carb tube w/ 10 mL water
2. Pass 900 mL sample through tube
3. Rinse tube w/10 mL water
4. Dry tube for 10 min.
5. Elute with 10 mL 80:20 methylene chloride: methanol containing .01% phosphoric acid

Minimal background from ENVI-Carb tube

Column: **polymeric-coated silica-based PAH specialty column, 20cm x 3mmID, 5µm particles**  
 Mobile Phase: gradient, A = water/0.05% H<sub>3</sub>PO<sub>4</sub>, B = acetonitrile  
 Temp.: 50°C  
 FlowRate: 0.5mL/min  
 Det.: photodiode array: peak width—0.053 min, sampling interval—0.320 sec, monitor 210nm & 225nm  
 Inj.: 10µL of extract (see Table 1)  
 (4-5 ppb each analyte in water)

| Time (min) | % B |
|------------|-----|
| 2.5        | 40  |
| 5.0        | 60  |
| 13.0       | 60  |
| 13.5       | 40  |

1. Tetrachloroterephthalic acid
2. Picloram
3. Dicamba
4. 2,4-D
5. 2,4,5-T
6. 2,4,5-TP
7. Dinoseb
8. Dacthal
9. Pentachlorophenol



Figures provided by A. Lichtman, Nassau County Dept. of Health, Hempstead, New York, USA.



## Acidic herbicide recovery using ENVI-Carb

Results: average recovery of 4 sample replicates spiked at 4-5 ug/L

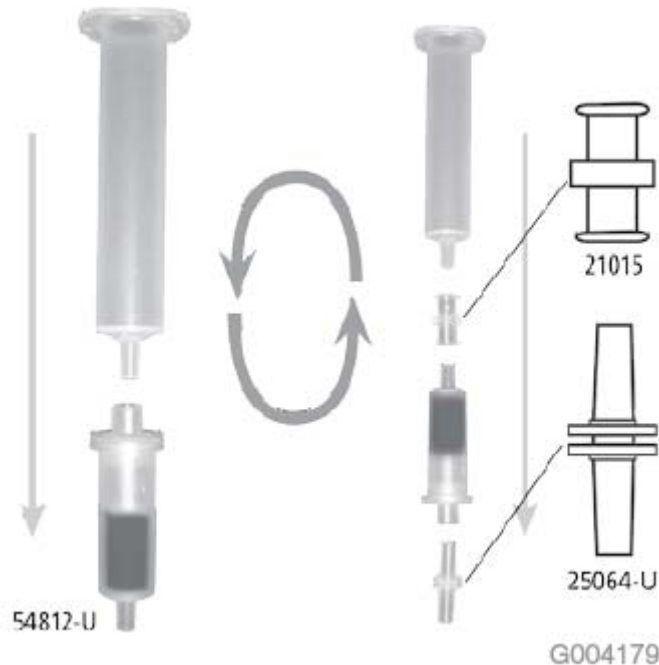
Lower recovery due to strong retention on carbon

| Analyte                      | % Recovery | Std. Dev. |
|------------------------------|------------|-----------|
| Tetrachloroterephthalic acid | 15.7       | -         |
| Picloram                     | 91.5       | 2.3       |
| Dicamba                      | 91         | 3.4       |
| 2,4-D                        | 88.3       | 7.1       |
| 2,4,5-T                      | 78.7       | 6.2       |
| 2,4,5,-TP                    | 84.7       | 9.9       |
| Dinoseb                      | 95.7       | 5.7       |
| Dachtal                      | 89.3       | 4.7       |
| Pentachlorophenol            | 69.5       | 7.6       |

- The smaller, more polar herbicides show good recoveries from ENVI-Carb.
- Tetrachloroerephthalic acid and pentachlorophenol are large with more planar structures. These are difficult to remove.

# Carbon SPE

## ENVI-Carb™ PLUS



### Spherical Carbon Molecular Sieve

- Extraction of highly polar compounds from water samples
- > 70% Absolute Recovery from 0.5 L drinking water (1-100 ng/mL)

### Procedure:

1. Condition w/ 10 mL MeOH & 10 mL DI water
2. Load up to 1 L sample
3. Reverse tube & elute w/ 4-5 mL MeOH in opposite direction

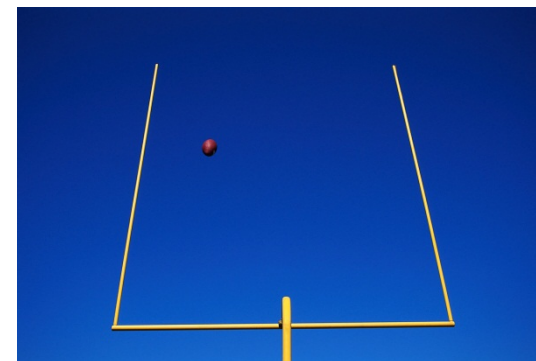
## Application: Analysis of Glycols In Water Using ENVI-Carb Plus

Current method requires direct aqueous injection (DAI)

- Water has a high vapor volume
  - Requires low injection volume and/or high inlet pressure to contain vapor cloud
- Poor peak shape
  - Especially early eluting compounds
  - Affects detection and quantitation
- Presence of matrix (such as salt) in a sample could easily foul the GC inlet and/or column.

# Analysis of Glycols

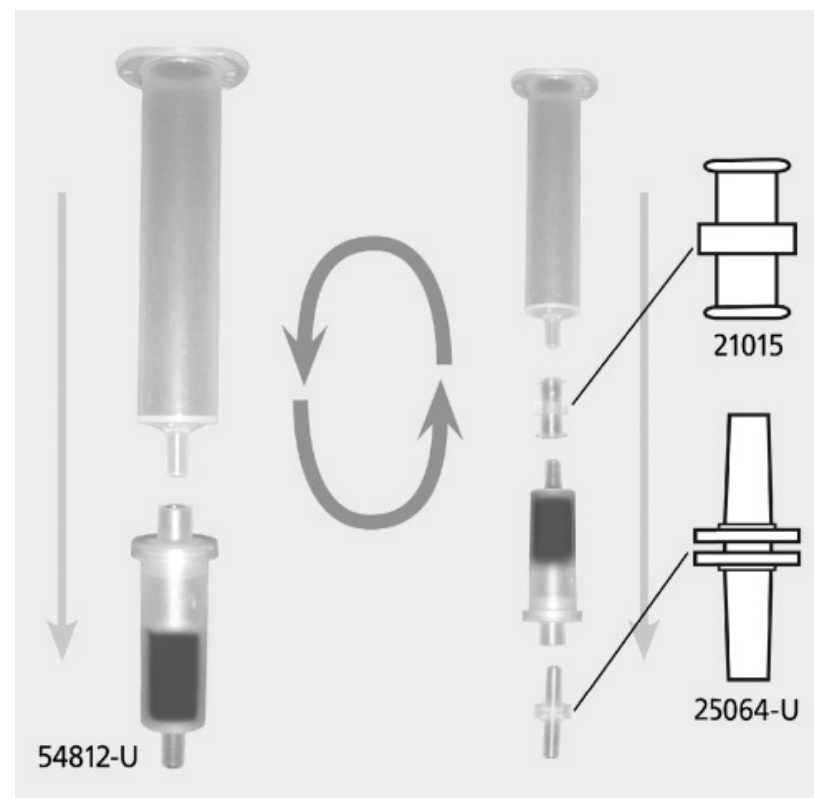
## Goals of New Method



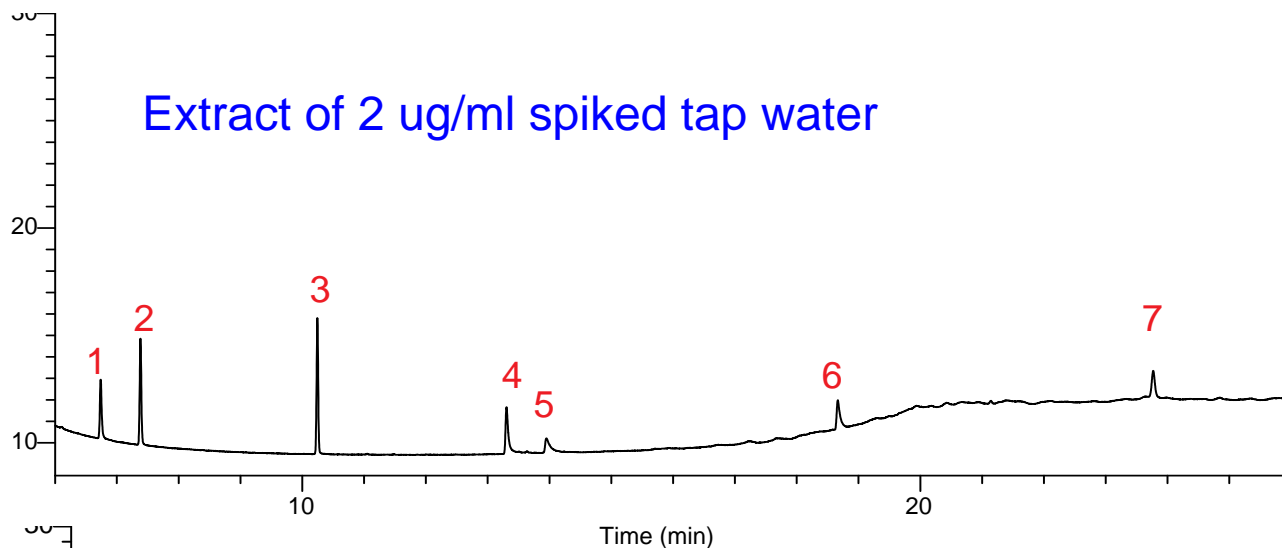
- Eliminate direct aqueous injection (DAI) of samples for glycol analysis.
- Lower detection and quantitation levels.
- Use solid phase extraction to extract glycols from water matrices.
  - Achieve elution with an organic solvent
  - Inject sample extract in organic solvent into GC

## Glycol SPE Extraction Method Using ENVI-Carb Plus

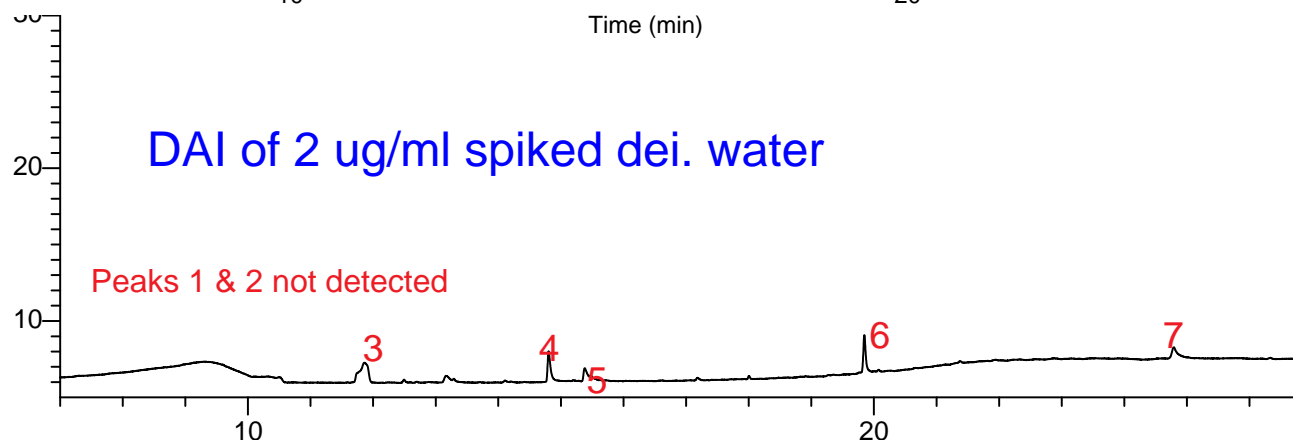
|                                    |   |
|------------------------------------|---|
| <b>Extraction Cartridge</b>        | ENVI-Carb™ Plus reversible cartridge, 1 ml/400 mg   |
| <b>Cartridge Conditioning</b>      | 1 ml methylene chloride , 2 x 2 ml aliquots methanol, 3 ml dei. water   |
| <b>Sample</b>                      | 5 mL tap water spiked at 10 ug/ml   |
| <b>Sample extraction</b>           | 5 ml sample, 5 mm Hg  |
| <b>Dry time</b>                    | 10 minutes, 10 mm Hg  |
| <b>Elution</b>                     | Cartridge in <b>forward</b> direction, 2 ml of methanol:methylene chloride, 80:20 (soak cartridge for 1 minutes prior to pulling through) |
| <b>Preparation for GC analysis</b> | Added methanol to bring sample to final volume of 2 ml and analyzed directly  |



# Extraction method vs. DAI



1. 2-methoxyethanol
2. 2-ethoxyethanol
3. 2-butoxyethanol
4. Propylene glycol
5. Ethylene glycol
6. Diethylene glycol
7. Triethylene glycol



**Column:** SPB-1000, 30 m x 0.25 mm I.D. x 0.25  $\mu$ m

**Oven:** 50 ° C (1 min. or 2.5 min.), 8 ° C/min. to 200 ° C (12 min.)

**Injector:** 220 ° C

**Carrier gas:** Helium, 1.5 ml/min, constant flow

**Detector:** FID, 220 ° C

**Injection:** 1  $\mu$ L, splitless

**Liner:** 4 mm ID, focus liner with taper (extracts) & 4 mm ID dual taper liner (DAI)

# Recovery and Reproducibility Evaluation

## Comparison of extraction method with DAI

Absolute recovery from tap water spiked at 2 ug/ml, n=7

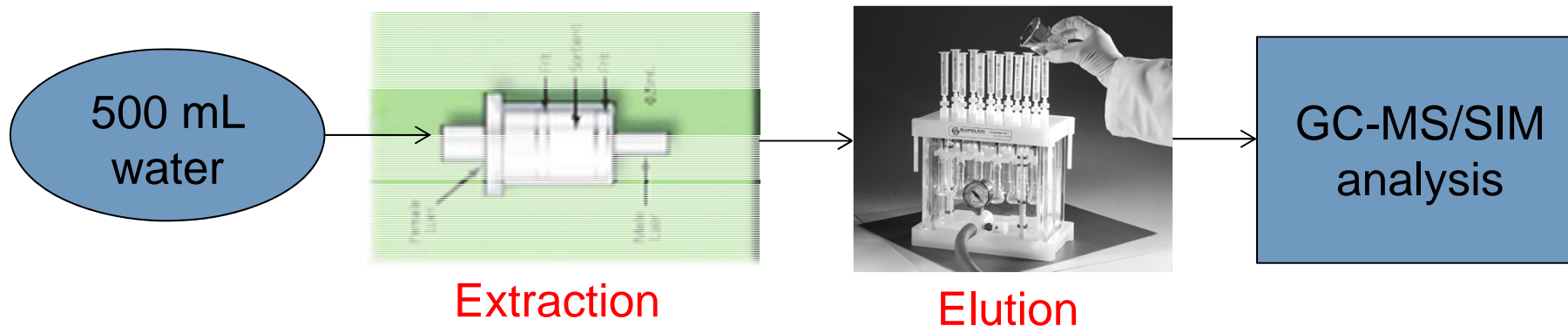
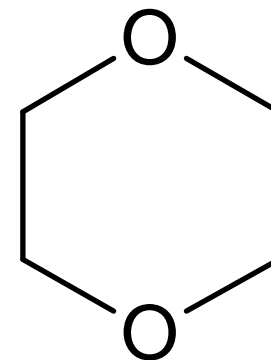
|                    | DAI             |          | Extraction      |          |
|--------------------|-----------------|----------|-----------------|----------|
|                    | Avg. % Recovery | %RSD n=7 | Avg. % Recovery | %RSD n=7 |
| 2-methoxyethanol   | Not detected    |          | 96%             | 3        |
| 2-ethoxyethanol    |                 |          | 88%             | 3        |
| 2-butoxyethanol    | 34%             | 30       | 94%             | 3        |
| Propylene glycol   | 106%            | 26       | 95%             | 6        |
| Ethylene glycol    | 114%            | 30       | 53%             | 16       |
| Diethylene glycol  | 56%             | 28       | 99%             | 6        |
| Triethylene glycol | 77%             | 26       | 98%             | 7        |

High RSDs

EG, smallest, most hydrophilic – difficult to retain on SPE

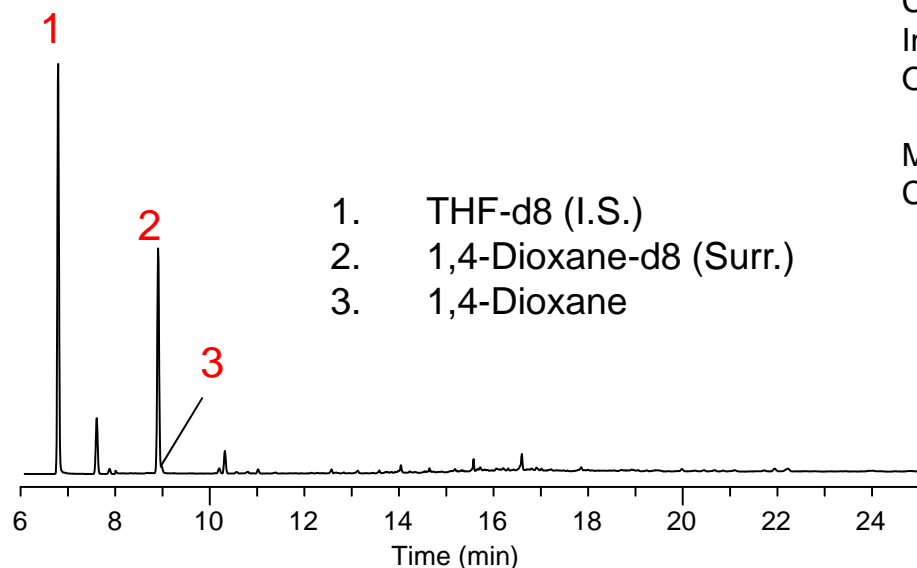
## Application: Extraction of 1,4-Dioxane from water using ENVI-Carb Plus

- US EPA Method 522 describes several different sorbents for extraction, including ENVI-Carb Plus.
- Flow through design of cartridge allows it to be connected directly to a continuous flow pump.
- ENVI-Carb Plus cartridges can be reversed (if necessary) for efficient elution of analytes.





# Analysis of 1,4-dioxane from water, extracted using ENVI-Carb Plus



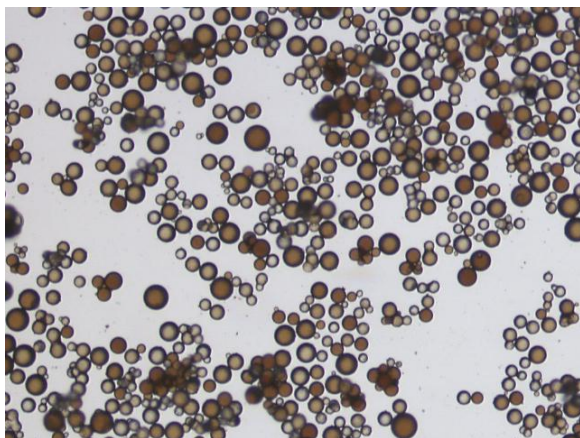
Column: SPB-624; 30 m x 0.25 mm I.D. x 1.4  $\mu$ m  
Inj. temp.: 200° C  
Oven: 30° C (1 min.), 13° C/min. to 90° C,  
20° C/min. to 200° C (15 min.)  
MS interface: 220° C  
Carrier: helium, 1 mL/min constant

|                    | 1,4-dioxane-d8<br>(surrogate) | 1,4-dioxane |
|--------------------|-------------------------------|-------------|
| Spike level (ug/L) | 10                            | 0.3         |
| Avg. % Recovery    | 102                           | 83          |
| %RSD, n=9          | 6                             | 4           |

# Supel<sup>TM</sup>-Select HLB

## Supel™- Select HLB

- Hydrophilic modified styrene-based polymer
- Can retain lipophilic and hydrophobic compounds
- Wide pH compatibility range (0-14)
- Imparts minimal extractables to samples
- Described in US EPA Method 1694 for the extraction of personal care and pharmaceutical compounds from water
- Very reproducible lot-to-lot performance



Optical microscope picture of  
Supel-Select HLB resin

# Extraction of Pharmaceutical & Personal Care Products (PPCPs) from Water using SupelSelect HLB

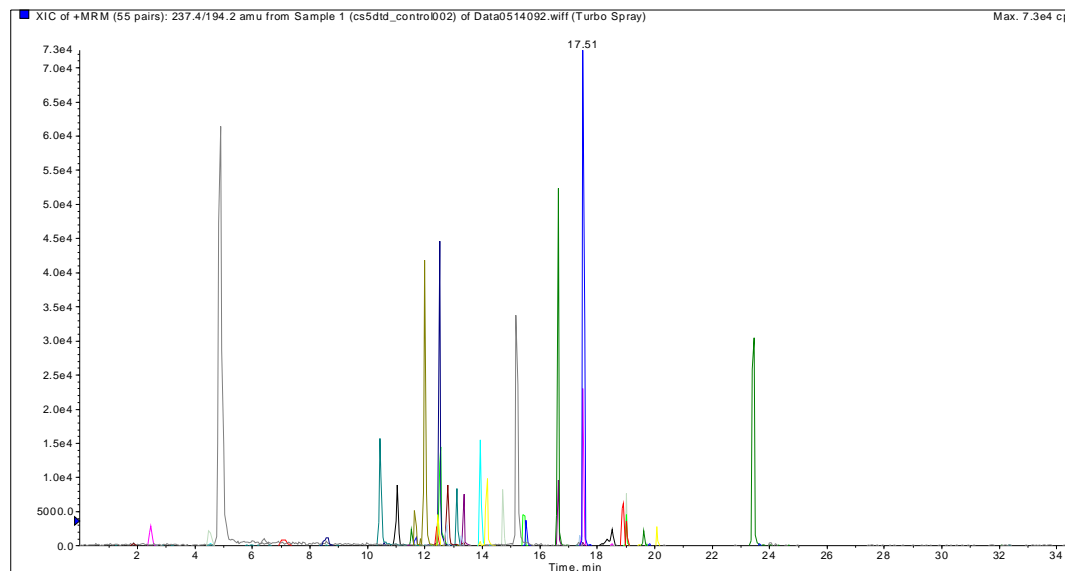
- US EPA Method 1694
- Method uses solid phase extraction (SPE) and LC-MS-MS for analysis of >70 PPCPs
- SPE used is hydrophilic/lipophilic balance (HLB)
  - Supel-Select HLB SPE

## Extraction – based on method 1694

|                      |  |
|----------------------|--|
| Extraction Cartridge | Supelco Select HLB SPE Tube, 500 mg/6 mL   |
| Sample               | 500 mL of drinking water spiked with Group 1 compounds, adjusted to pH=4 with 6M HCl |
| Tube Conditioning    | 20 mL of methanol, 6 mL of water, 6 mL of water at pH 2 (with 6 M HCl)               |
| Sample extraction    | 10 mL/min through tube   |
| Dry time             | 5 min  |
| Elution              | 12 mL, 50:50 methanol:acetonitrile   |
| Dry down             | 40 ° C, under nitrogen stream  |
| Reconstitution       | To final volume of 2 mL using mobile phase   |



# LC-MS/MS Analysis



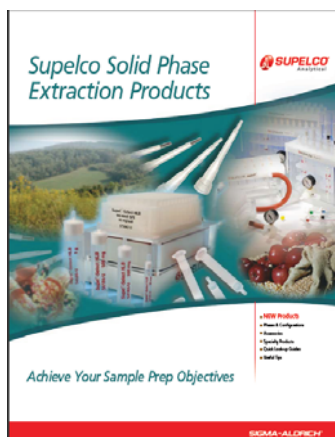
|                 |  |               |            |            |
|-----------------|--|---------------|------------|------------|
| column:         | Ascentis Express C18, 10cm x 2.1 mm, I.D., 2.7 $\mu$ m (53823-U) |               |            |            |
| mobile phase A: | 0.1% formic acid and 0.1 % ammonium formate                      |               |            |            |
| mobile phase B: | 50:50 methanol:acetonitrile                                      |               |            |            |
| temp.:          | 40 ° C   |               |            |            |
| det.:           | ESI (+), MS/MS   |               |            |            |
| injection:      | 5 $\mu$ L  |               |            |            |
| sample:         | CS-5 Concentrations from EPA method 1694                         |               |            |            |
| gradient:       | <u>min</u>   | <u>mL/min</u> | <u>% A</u> | <u>% B</u> |
|                 | 0  | 0.15          | 95         | 5          |
|                 | 4  | 0.25          | 95         | 5          |
|                 | 22.5   | 0.30          | 12         | 88         |
|                 | 23   | 0.30          | 0          | 100        |
|                 | 26   | 0.30          | 0          | 100        |
|                 | 26.5   | 0.15          | 95         | 5          |

Fused core column for efficient, fast analysis

## Recoveries of some PPCPs from spiked water samples - extracted using Supel-Select HLB SPE

| Compound             | MRM Transition | SPE Spiking Level (ug/L) | % Recovery |
|----------------------|----------------|--------------------------|------------|
| Azithromycin         | 749.9-591.6    | 1                        | 95         |
| Caffeine             | 195.0-138.0    | 10                       | 129        |
| Carbadox             | 263.2-231.2    | 1                        | 123        |
| Ciprofloxacin        | 332.2-314.2    | 3,5                      | 105        |
| Clinafloxacin        | 366.3-348.1    | 4                        | 48         |
| Digoxigenin          | 391.2-355.2    | 4                        | 128        |
| Digoxin              | 781.5-113.1    | 10                       | 136        |
| 1,7-Dimethylxanthine | 181.2-124.0    | 100                      | 86         |
| Diphenhydramine      | 256.8-168.1    | 0,4                      | 122        |
| Enrofloxacin         | 360.0-316.0    | 2                        | 83         |
| Lomefloxacin         | 352.2-308.1    | 2                        | 106        |
| Miconazole           | 417.0-161.0    | 1                        | 74         |
| Norfloxacin          | 320.0-302.0    | 10                       | 75         |
| Ofloxacin            | 362.2-318.0    | 1                        | 83         |
| Ormetoprim           | 275.3-259.1    | 0,4                      | 128        |
| Oxolinic acid        | 244.1-216.1    | 0,4                      | 77         |
| Sarafloxacin         | 386.0-299.0    | 9,12                     | 100        |
| Thiabendazole        | 202.1-175.1    | 1                        | 119        |

# Key SPE Literature



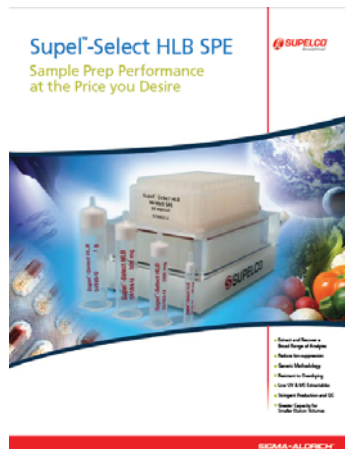
**SPE Brochure  
T402150D**



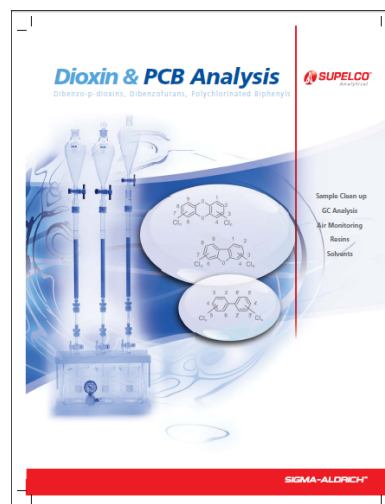
**SPE Methodology Wall  
Poster T409088**



**Empore SPE Flyer  
T407075**



**Supel Select HLB  
Brochure T408148**



**Dioxin & PCB Analysis**



**Environmental Sampling &  
Analysis Guide T412043**

**Thank you!**



## Associated Supelco Publications

1. *Extract Polynuclear Aromatic Hydrocarbons from Water, Using Solid Phase Extraction Disks: Application Note 54 (T394054A).*
2. *Extraction of Paraquat and Diquat from Water, Using ENVI™-8 DSK Solid Phase Extraction Disks: Application Note 60 (T394060A).*
3. *ENVI™-18 SPE Tube Ensures Low Background from Monitoring Organic Compounds in Drinking Water by EPA Method 525: Application Note 65 (T395065A).*
4. *Extract Nonvolatile Pesticides from Drinking Water, Using a Graphitized Carbon Adsorbent: Application Note 27 (T394027B)*
5. *Solid Phase Extraction/HPLC Analysis of Acidic Herbicides in Drinking Water. Application Note 100 (T396100).*
6. Stenerson, K.K.; Sidisky, L.M.; Betz, W.R.; Keeler, M.J.; McCoy, M.; Brown, J.. *The Extraction of Glycols from Water Using ENVI-Carb™ Plus Solid Phase Extraction Cartridges (T410135)*
7. Santasania, C.T.; Stenerson, K.K.; Aurand, C.; Trinh, A.; Shirey R.E. *Using Bonded Silica Solid Phase Microextraction Fibers as a Screening Tool for Pharmaceuticals and Personal Care Products in Drinking Water (T409139)*

## References

1. Methods for the Determination of Organic Compounds in Drinking Water Supplement 1; EPA/600/R/4-90/020
2. Methods for the Determination of Organic and Inorganic Compounds in Drinking Water Volume 1; EPA 815-R-00-014
3. Munch, J.W.; Grimmett, P.E. Method 522 *Determination of 1,4-Dioxane in Drinking Water by Solid Phase Extraction (SPE) and Gas Chromatography/Mass Spectrometry (GC/MS) With Selected Ion Monitoring (SIM)*. Ver. 1; EPA /600/R-08/101; National Exposure Research Laboratory Office of Research and Development U.S. Environmental Protection Agency: Cincinnati, OH, Sept. 2008.
4. Method 1694: *Pharmaceuticals and Personal Care Products in Water, Soil, Sediment, and Biosolids by HPLC/MS/MS*. EPA-821-R-08-002; U.S. Environmental Protection Agency Office of Water Office of Science and Technology Engineering and Analysis Division (4303T): Washington, D.C., Dec. 2007.